



# High-Sensitivity Troponin I (TnIH)

Current Revision and Date <sup>a</sup>	Rev. 07, 2022-10	
Product Name	Atellica IM High-Sensitivity Troponin I (TnIH)	REF 10997840 (100 tests)
		REF 10997841 (500 tests)
Abbreviated Product Name	Atellica IM TnIH	
Test Name/ID	TnlH	
Systems	Atellica CI Analyzer	
Materials Required but Not Provided	Atellica IM APW3	REF 10998580
Optional Materials	Atellica IM Multi-Diluent 11	REF 10995642 (2-pack)
	Atellica IM TnIH MCM	REF 10997842
Specimen Types	Serum, lithium-heparin plasma	
Sample Volume	100 μL	
Measuring Interval	2.50-25,000.00 pg/mL (ng/L)	

<sup>&</sup>lt;sup>a</sup> A vertical bar in the page margin indicates technical content that differs from the previous version.

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### Intended Use

The Atellica® IM High-Sensitivity Troponin I (TnIH) assay is for *in vitro* diagnostic use in the quantitative measurement of cardiac troponin I in human serum or plasma (lithium heparin) using the Atellica® CI Analyzer. The assay can be used to aid in the diagnosis of acute myocardial infarction (AMI).

# **Summary and Explanation**

Troponin I (TnI) exists in 3 distinct isoforms: cardiac muscle, slow-twitch skeletal muscle, and fast-twitch skeletal muscle. Each isoform is encoded by a distinct gene, each with a unique amino acid sequence, leading to a 40% dissimilarity among isoforms. 1–4

Cardiac troponin I (cTnI) is an inhibitory protein of the troponin-tropomyosin complex. cTnI is the only TnI isotype present in the myocardium and is not expressed during any developmental stage in skeletal muscle.<sup>2,5,6</sup> cTnI has a molecular weight of 24,000 daltons.<sup>7</sup>

The cardiac form of TnI is further unique in that it has 31 additional amino acid residues on its N-terminal, not present in the skeletal forms, which allows for specific monoclonal antibody development.<sup>7</sup> The cardiac specificity of this isoform improves the accuracy of detection of cardiac muscle ischemia in patients with acute or chronic skeletal muscle injury and possible concomitant myocardial injury, and is the basis for its selection as a cardiac marker in the diagnosis of AMI.<sup>1,3–5,7,8</sup>

The Global MI Task Force's third version of the universal definition of myocardial infarction defined AMI as evidence of myocardial necrosis in a clinical setting consistent with acute myocardial ischemia. Under these circumstances, the following criterion meets the diagnosis of AMI:

Detection of a rise and/or fall of cardiac biomarker values (preferably cardiac troponin) with at least 1 value above the 99th percentile upper reference limit (URL) and with at least 1 of the following conditions:

- Symptoms of ischemia.
- New or presumed new significant ST-segment-T wave (ST-T) changes or new left bundle branch block (LBB).
- Development of pathological Q waves in the electrocardiogram (EKG).
- Imaging evidence of new loss of viable myocardium, or new regional wall motion abnormality.
- Identification of an intracoronary thrombus by angiography or autopsy.

# **Definition of a High-Sensitivity Assay**

The International Federation of Clinical Chemistry (IFCC) Task Force on Clinical Applications of Cardiac Bio-Markers defines a troponin assay as a high-sensitivity assay if it meets the following criteria:<sup>10</sup>

- Total imprecision (CV) at the 99th percentile value should be at or below 10%.
- Measurable concentrations should be attainable at concentrations above the limit of detection (LoD) in at least 50% of healthy individuals.

Troponin values must be used in the context of the patient clinical presentation. Serial sampling is recommended to detect the temporal rise and fall of troponin levels characteristic of AMI. The demonstration of a temporal rise and fall in troponin is needed to distinguish AMI from troponin elevations associated with non-AMI conditions, such as renal failure, arrhythmias, pulmonary embolism, chronic renal disease, myocarditis, and cardiotoxicity.<sup>9,11–14</sup>

# **Principles of the Procedure**

The Atellica IM TnIH is a 3-site sandwich immunoassay using direct chemiluminometric technology. The Solid Phase reagent is magnetic latex particles conjugated with streptavidin with 2 bound biotinylated capture monoclonal antibodies each recognizing a unique cTnI epitope.

The Lite Reagent comprises a conjugate whose architecture consists of a proprietary acridinium ester and a recombinant anti-human cTnI sheep Fab covalently attached to bovine serum albumin (BSA) for chemiluminescent detection.

A direct relationship exists between the amount of troponin I present in the patient sample and the amount of relative light units (RLUs) detected by the system.

# Reagents

Material Description	Storage	Stability <sup>a</sup>
Atellica IM TnIH ReadyPack® primary reagent pack Lite Reagent	Unopened at 2–8°C	Until expiration date on product
8.0 mL/reagent pack Bovine serum albumin (BSA) conjugated to a recombinant monoclonal (sheep) Fab anti-human cTnl (~0.2–0.4 µg/mL) labeled with acridinium ester in HEPES buffer; stabilizers; preservatives  Solid Phase 13.0 mL/reagent pack Streptavidin-coated magnetic latex particles (0.45 mg/mL) with 2 biotinylated (mouse and sheep) monoclonal anti-troponin I antibodies in buffer; stabilizers; preservatives	Onboard	90 days
Atellica IM TnIH CAL L  1.0 mL/vial	Unopened at 2–8°C	Until expiration date on product
HEPES buffer; bovine serum albumin (BSA); surfactants; preservatives	Opened at 2–8°C	4 hours
	Opened at ≤ -20°C	30 days; thaw 1 time
	Onboard at room temperature	4 hours
Atellica IM TnIH CAL H 1.0 mL/vial; lyophilized	Lyophilized at 2–8°C	Until expiration date on product
After reconstitution, human serum; human cTnI; preservatives	Reconstituted at 2–8°C	4 hours
	Reconstituted at ≤ -20°C	30 days; thaw 1 time
	Onboard at room temperature	4 hours
Atellica IM APW3 ReadyPack ancillary reagent pack <sup>b</sup> 25.0 mL/pack	Unopened at 2–8°C	Until expiration date on product
Phosphate-buffered saline; sodium azide (< 0.1%); surfactant	Onboard	28 days
Atellica IM Multi-Diluent 11 ReadyPack ancillary reagent pack <sup>c</sup> 5.0 mL/pack	Unopened at 2–8°C	Until expiration date on product
Tris buffer; goat serum; protein stabilizers; preservatives	Onboard	28 days

- <sup>a</sup> Refer to Storage and Stability.
- <sup>b</sup> Refer to Materials Required but Not Provided.
- c Refer to Optional Materials.

# **Warnings and Precautions**

For in vitro diagnostic use.

For Professional Use.

#### **CAUTION**

Federal (USA) law restricts this device to sale by or on the order of a licensed healthcare professional.

Safety data sheets (SDS) available on siemens-healthineers.com.



#### **CAUTION POTENTIAL BIOHAZARD**

Contains human source material. Each donation of human blood or blood component was tested by FDA-approved methods for the presence of antibodies to human immunodeficiency virus type 1 (HIV-1) and type 2 (HIV-2), as well as for hepatitis B surface antigen (HBsAg) and antibody to hepatitis C virus (HCV). The test results were negative (not repeatedly reactive). No test offers complete assurance that these or other infectious agents are absent; this material should be handled using good laboratory practices and universal precautions.<sup>15–17</sup>

#### **CAUTION**

This device contains material of animal origin and should be handled as a potential carrier and transmitter of disease.

Contains sodium azide as a preservative. Sodium azide can react with copper or lead plumbing to form explosive metal azides. On disposal, flush reagents with a large volume of water to prevent buildup of azides. Disposal into drain systems must be in compliance with prevailing regulatory requirements.

Dispose of hazardous or biologically contaminated materials according to the practices of your institution. Discard all materials in a safe and acceptable manner and in compliance with prevailing regulatory requirements.

**Note** For information about reagent preparation, refer to *Preparing the Reagents* in the *Procedure* section.

**Note** For information about calibrator preparation, refer to *Preparing the Calibrators*.

## Storage and Stability

Store reagents in an upright position. Protect the product from heat and light sources. Unopened reagents are stable until the expiration date on the product when stored at  $2-8^{\circ}$ C.

Store Atellica IM TnIH CAL L in an upright position. Unopened calibrator is stable until the expiration date on the product when stored at  $2-8^{\circ}$ C. Opened calibrator is stable for 4 hours at  $2-8^{\circ}$ C. Freeze opened product at  $\leq -20^{\circ}$ C for up to 30 days; thaw 1 time. Calibrators are stable for 4 hours at room temperature.

Store Atellica IM TnIH CAL H in an upright position. Lyophilized calibrator is stable until the expiration date on the product when stored at  $2-8^{\circ}$ C. Reconstituted calibrator is stable for 4 hours at  $2-8^{\circ}$ C. Freeze reconstituted product at  $\leq -20^{\circ}$ C for up to 30 days; thaw 1 time. Reconstituted calibrators are stable for 4 hours at room temperature.

Store Atellica IM APW3 in an upright position. Unopened Atellica IM APW3 is stable until the expiration date on the product when stored at  $2-8^{\circ}$ C.

Store Atellica IM Multi-Diluent 11 in an upright position. Unopened Atellica IM Multi-Diluent 11 is stable until the expiration date on the product when stored at  $2-8^{\circ}$ C.

Do not use products beyond the expiration date printed on the product labeling.

# **Onboard Stability**

Discard products at the end of the onboard stability interval. Do not use products beyond the expiration date printed on the product labeling.

For information about product onboard stability, refer to Reagents.

# **Specimen Collection and Handling**

Serum and plasma (lithium heparin) are the recommended sample types for this assay.

## **Collecting the Specimen**

• Observe universal precautions when collecting specimens. Handle all specimens as if they are capable of transmitting disease.<sup>17</sup>

- Follow recommended procedures for collection of diagnostic blood specimens by venipuncture.<sup>18</sup>
- Follow the instructions provided with your specimen collection device for use and processing.<sup>19</sup>
- Allow blood specimens to clot completely before centrifugation.<sup>16</sup>
- Keep tubes capped at all times. 16
- The use of a single sample type (either lithium-heparin plasma or serum) is recommended for troponin analysis when collecting serial samples from the same patient.
- For serum specimens, complete clot formation should take place before centrifugation.
   Serum should be physically separated from cells as soon as possible from the time of collection.<sup>16</sup>
- Samples must be free of fibrin or other particulate matter. The presence of fibrin, red blood cells, or suspended particles may lead to inaccurate results. Serum samples that contain suspended fibrin particles or erythrocyte stroma must be re-centrifuged before testing.
- If clotting time is increased due to thrombolytic or anticoagulant therapy, the use of plasma specimens will allow for faster sample processing and reduce the risk of microclots, fibrin or particulate matter.
- For plasma specimens, avoid transferring white blood cells or platelets from the layer located just above the red blood cells.
- If a fixed angle rotor is used for centrifugation, care should be taken to avoid resuspending cellular material (platelets) upon removal from the centrifuge.

## **Storing the Specimen**

- Samples are stable up to 8 hours when tightly capped and stored at room temperature.
- Samples are stable up to 24 hours when tightly capped and stored at 2–8°C.
- Samples can be frozen at  $\leq$  -20°C for up to 40 days. Do not store in a frost-free freezer.
- Samples can be frozen at  $\leq$  -70°C for up to 1 year.
- Freeze samples only once and mix thoroughly after thawing.

#### **CAUTION**

Thoroughly mix thawed samples and centrifuge them before using. <sup>16</sup> Collect the supernatant into a clean vial.

The handling and storage information provided here is based on data or references maintained by the manufacturer. It is the responsibility of the individual laboratory to use all available references and/or its own studies when establishing alternate stability criteria to meet specific needs.

# Transporting the Specimen

Ship samples frozen.

Package and label specimens for shipment in compliance with applicable federal and international regulations covering the transport of clinical specimens and etiological agents.

## **Preparing the Samples**

This assay requires  $100 \, \mu L$  of sample for a single determination. This volume does not include the unusable volume in the sample container or the additional volume required when performing duplicates or other tests on the same sample. For information about determining the minimum required volume, refer to the online help.

The sample volume required to perform onboard dilution differs from the sample volume required to perform a single determination. Refer to *Dilutions*.

**Note** Do not use specimens with apparent contamination.

Before placing samples on the system, ensure that samples are free of:

- Bubbles or foam.
- Fibrin or other particulate matter.

**Note** Remove particulates by centrifugation according to CLSI guidance and the collection device manufacturer's recommendations. 16

**Note** For a complete list of appropriate sample containers, refer to the online help.

### **Procedure**

#### **Materials Provided**

The following materials are provided:

REF	Contents	Number of Tests
10997840	1 ReadyPack primary reagent pack containing Atellica IM TnIH Lite Reagent and Solid Phase TnIH master curve and test definition MCTDEF  1 vial Atellica IM TnIH CAL low calibrator CAL L  1 vial Atellica IM TnIH CAL high calibrator CAL H  Atellica IM TnIH CAL calibrator lot-specific value sheet CAL LOT VAL	100
10997841	5 ReadyPack primary reagent packs containing Atellica IM TnIH Lite Reagent and Solid Phase TnIH master curve and test definition MCTDEF  2 vials Atellica IM TnIH CAL low calibrator CAL L  2 vials Atellica IM TnIH CAL high calibrator CAL H  Atellica IM TnIH CAL calibrator lot-specific value sheet CAL LOT VAL	500

# **Materials Required but Not Provided**

The following materials are required to perform this assay, but are not provided:

R	EF	Description	
		Atellica CI Analyzer <sup>a</sup>	
10	0998580	Atellica IM APW3 (probe wash)	2 ReadyPack ancillary reagent packs containing 25.0 mL/pack WASH

<sup>&</sup>lt;sup>a</sup> Additional system fluids are required to operate the system. For system fluid instructions for use, refer to the Document Library.

### **Optional Materials**

The following materials may be used to perform this assay, but are not provided:

REF	Description	
10995642	Atellica IM Multi-Diluent 11 (diluent)	2 ReadyPack ancillary reagent packs containing 5.0 mL/pack DIL
10997842	Atellica IM TnIH MCM (master curve material)	5 x 1.0 mL levels of master curve material MCM

## **Assay Procedure**

The system automatically performs the following steps:

- 1. Dispenses 100  $\mu$ L of sample into a cuvette.
- 2. Dispenses 130  $\mu$ L of Solid Phase and 80  $\mu$ L of Lite Reagent, then incubates for 8 minutes at 37°C.
- 3. Separates, aspirates, then washes the cuvette with Atellica IM Wash.
- 4. Dispenses 300  $\mu$ L each of Atellica IM Acid and Atellica IM Base to initiate the chemiluminescent reaction.
- 5. Reports results.

## **Preparing the Reagents**

All reagents are liquid and ready to use. Before loading primary reagent packs onto the system, mix them by hand and visually inspect the bottom of the reagent pack to ensure that all particles are resuspended. For information about preparing the reagents for use, refer to the online help.

# **Preparing the System**

Ensure that the system has sufficient reagent packs loaded in the reagent compartment. The system automatically mixes reagent packs to maintain homogeneous suspension of the reagents. For information about loading reagent packs, refer to the online help.

For automated dilutions, ensure that Atellica IM Multi-Diluent 11 is loaded on the system.

#### **Master Curve Definition**

Before initiating calibration on each new lot of reagent, load the assay master curve and test definition values by scanning the MCTDEF 2D barcodes. For loading instructions, refer to the online help.

# Performing Calibration

For calibration of the Atellica IM TnIH assay, use the calibrators provided with each kit.

## **Calibration Frequency**

Perform a calibration if one or more of the following conditions exist:

- When changing lot numbers of primary reagent packs.
- At the end of the lot calibration interval, for a specified lot of calibrated reagent on the system.
- At the end of the pack calibration interval, for calibrated reagent packs on the system.

- When indicated by quality control results.
- After major maintenance or service, if indicated by quality control results.

At the end of the onboard stability interval, replace the reagent pack on the system with a new reagent pack. Recalibration is not required, unless the lot calibration interval is exceeded.

Stability Interval	Days
Lot Calibration	47
Pack Calibration	35
Reagent Onboard Stability	90

For information about lot calibration and pack calibration intervals, refer to the online help.

Follow government regulations or accreditation requirements for calibration frequency. Individual laboratory quality control programs and procedures may require more frequent calibration.

### **Preparing the Calibrators**

#### Low Calibrator

The Atellica IM TnIH CAL L is liquid and ready to use. Gently mix and invert the vials to ensure homogeneity of the material.



DO NOT ADD WATER TO THE LOW CALIBRATOR.

#### **High Calibrator**

Prepare the Atellica IM TnIH CAL H using the following steps:

1. Add 1.00 mL of special reagent water into the vial using a class A volumetric pipet or an equivalent pipet. Replace cap.

Note For information about special reagent water requirements, refer to the online help.

- 2. Let the vial stand for 15–20 minutes at room temperature to allow the lyophilized material to dissolve.
- 3. Gently mix and invert the vial to ensure homogeneity of the material.

**Note** For extended storage, aliquot the low and high calibrators into cryovials and seal tightly. Store material according to stability limits specified in *Storage and Stability*. Do not store in a frost-free freezer.

Before using frozen calibrators, allow the material to completely thaw. Gently mix and invert the vials to ensure homogeneity of the material. Use immediately and discard any remaining material.

**Note** Use calibrators within the stability limits specified in *Storage and Stability* and discard any remaining material.

#### **Calibration Procedure**

The required sample volume for testing depends on several factors. For information about sample volume requirements, refer to the online help.

Use the following lot-specific materials to perform calibration:

• For the master curve and assay test definitions, refer to the lot-specific master curve and test definition sheet MCTDEF provided with the assay reagents.

- Calibrators provided in an assay kit must only be used with reagents from that assay kit lot. Do not use calibrators from one assay kit with reagents from a different assay kit lot.
- For the calibrator definitions, refer to the lot-specific value sheet [CAL LOT VAL] provided with the calibrator materials.
- Generate lot-specific barcode labels to use with the calibrator samples.

For instructions about how to perform the calibration procedure, refer to the online help.

## **Performing Quality Control**

For quality control of the Atellica IM TnIH assay, use an appropriate quality control material of known analyte concentration with at least 2 levels at least once during each day that samples are analyzed. Use the quality control material in accordance with the quality control instructions for use.

A satisfactory level of performance is achieved when the analyte values obtained are within the expected control interval for the system or within your interval, as determined by an appropriate internal laboratory quality control scheme. Follow your laboratory's quality control procedures if the results obtained do not fall within the acceptable limits. For information about entering quality control definitions, refer to the online help.

Follow government regulations or accreditation requirements for quality control frequency. Individual laboratory quality control programs and procedures may require more frequent quality control testing.

Test quality control samples after a successful calibration.

### **Taking Corrective Action**

If the quality control results do not fall within the assigned values, do not report results. Perform corrective actions in accordance with established laboratory protocol. For suggested protocol, refer to the online help.

### Results

#### Calculation of Results

The system determines the result using the calculation scheme described in the online help. The system reports results in pg/mL (common units) or ng/L (SI units), depending on the units defined when setting up the assay.

Conversion formula: 1.0 pg/mL (common units) = 1.0 ng/L (SI units)

For information about results outside the specified measuring interval, refer to *Measuring Interval*.

#### **Dilutions**

The measuring interval for serum and plasma is 2.50–25,000.00 pg/mL (ng/L). For information about dilution options, refer to the online help.

Patient samples with cTnI levels > 25,000.00 pg/mL (ng/L) can be diluted and retested to obtain quantitative results. Patient samples with cTnI levels  $\leq$  25,000.00 pg/mL (ng/L) should not be diluted.

For automated dilutions, ensure that Atellica IM Multi-Diluent 11 is loaded in the reagent compartment. Ensure that sufficient sample volume is available to perform the dilution and that the appropriate dilution factor is selected when scheduling the test, as indicated in the table below.

For automatic dilutions, enter a dilution setpoint  $\leq$  25,000.00 pg/mL (ng/L).

Sample	Dilution	Sample Volume (μL)
Serum and plasma	1:2	100
Serum and plasma	1:5	60

## Interpretation of Results

Results of this assay should always be interpreted in conjunction with the patient's medical history, clinical presentation, and other findings.

### Limitations

The following information pertains to limitations of the assay:

- The use of a single sample type (either lithium heparin or serum) is recommended for troponin analysis when collecting serial samples from the same patient.
- If clotting time is increased due to thrombolytic or anticoagulant therapy, using serum samples may increase the risk of micro-clots, fibrin, or particulate matter. Lithium-heparin plasma is the preferred sample type for patients undergoing anticoagulant therapy.
- Samples from patients receiving preparations of mouse monoclonal antibodies for therapy or diagnosis may contain human anti-mouse antibodies (HAMA). Such samples may show either falsely elevated or falsely depressed values when tested with this method.<sup>20</sup>
- Specimens from some individuals with pathologically high gamma globulin levels may demonstrate depressed troponin values. This may be due to the presence of cardiac troponin-specific autoantibodies.<sup>21</sup> Additional information may be required for diagnosis.
- Patient samples may contain heterophilic antibodies that could react in immunoassays to give falsely elevated or depressed results. This assay is designed to minimize interference from heterophilic antibodies.<sup>22,23</sup>

# **Expected Values**

Expected values were established using the Atellica IM Analyzer and verified on the Atellica CI Analyzer in accordance with CLSI guideline EP28-A3c.<sup>24</sup> The reagents used on the Atellica CI Analyzer are the same as those used on the Atellica IM Analyzer.

Serum and lithium-heparin plasma specimens were collected from 2007 apparently healthy individuals from the United States who ranged in age from 22–91 years of age. Each specimen was frozen, thawed, and assayed once. The 99th percentile values were determined using the non-parametric statistical method described in CLSI Document EP28-A3c.<sup>24</sup> Two female subjects had troponin values of approximately 300 pg/mL (ng/L) and 5000 pg/mL (ng/L), and were considered to be outliers. These results were not included in the 99th percentile determination.

The 99th percentile values determined for lithium-heparin plasma (female, male, and combined), and for serum (female, male, and combined) are shown in the following table. The 90% confidence intervals demonstrate that there is no statistical basis for using separate 99th percentile values based on gender or sample type.

The combined gender and the more commonly used sample type of lithium-heparin plasma were used to determine the overall observed 99th percentile of 45.20 pg/mL (ng/L). In the IFCC-recommended reporting format (whole numbers), the 99th percentile is 45 pg/mL (ng/L).

Sample Type	Gender	N	99th Percentile <sup>a</sup> (pg/mL; ng/L)	90% CI <sup>b</sup> (pg/mL; ng/L)
Lithium heparin	Female	1007	34.11	27.36–66.23
	Male	1000	53.48	38.73-80.22
	Combined	2007	45.20	33.21–64.30
Serum	Female	1007	38.64	28.58-72.36
	Male	994	53.53	33.77–78.03
	Combined	2001	45.43	35.47-63.63

<sup>&</sup>lt;sup>a</sup> IFCC Task Force on Clinical Applications of Cardiac Bio-Markers recommends that troponin values be reported as whole numbers.<sup>10</sup>

As with all *in vitro* diagnostic assays, each laboratory should establish its own diagnostic cutoff value, which reflects criteria for AMI diagnosis at their institution and is representative of specific populations.

### **Performance Characteristics**

The reagents used on the Atellica CI Analyzer are the same as those used on the Atellica IM Analyzer. Some performance characteristics were established with the Atellica IM Analyzer.

## **Measuring Interval**

The Atellica IM TnIH assay is linear from 2.50–25,000.00 pg/mL (ng/L). The lower limit of the measuring interval is defined by the LoQ. Report patient results below the measuring interval as < 2.50 pg/mL (ng/L). When sample results exceed the measuring interval, refer to *Dilutions*.

# **Specificity**

The Atellica IM TnIH assay shows high specificity for cTnI. The following compounds were added at the concentrations indicated to a serum or lithium-heparin plasma sample with a known cTnI concentration. Atellica IM TnIH assay results from the spiked samples were compared with those of unspiked control samples. Percent cross-reactivity was determined in accordance with CLSI Document EP07-A2<sup>25</sup> and is calculated as:

% cross-reactivity =  $\frac{\text{(concentration of spiked sample - concentration of unspiked sample)}}{\text{concentration of compound}} \times 100$ 

Cross-reactant	Amount (ng/mL)	Cross-reactivity (%)
Cardiac troponin T <sup>a</sup>	1000	NDb
Skeletal troponin I	1000	ND
Tropomyosin	1000	ND
Actin	1000	ND
Troponin C	1000	ND
Myosin light chain	1000	ND

b Confidence interval.

Cross-reactant	Amount (ng/mL)	Cross-reactivity (%)
Myoglobin	1000	ND
СК-МВ	1000	ND

Human recombinant.

Results were established using the Atellica IM Analyzer. Assay results obtained at individual laboratories may vary from the data presented.

## **Detection Capability**

Limit of Blank (LoB)	0.50 pg/mL (ng/L)
Limit of Detection (LoD)	1.60 pg/mL (ng/L)
Limit of Quantitation (LoQ)	2.50 pg/mL (ng/L)

The LoB corresponds to the highest measurement result likely to be observed for a blank sample with a probability of 95%.

The LoD corresponds to the lowest concentration of cTNI that can be detected with a probability of 95%.

The LoQ corresponds to the lowest amount of analyte in a sample at which the within laboratory CV is 20%.

Detection capability was determined in accordance with CLSI Document EP17-A2.<sup>26</sup>

## **High-Sensitivity Determination**

The Atellica IM TnIH assay meets the IFCC Task Force on Clinical Applications of Cardiac Bio-Markers' definition of a high-sensitivity troponin assay. 10

- 1. Total imprecision (CV) at the 99th percentile value of 45.20 pg/mL (ng/L) is below 10%.
- 2. Greater than 50% of measurements from individuals in the healthy patient population used to determine the 99th percentile value were above the LoD of 1.60 pg/mL (ng/L).

### **Clinical Performance**

A prospective study was performed to assess diagnostic accuracy for subjects in both serum and lithium-heparin plasma sample types. Specimens were collected at 29 emergency departments across the United States, from subjects presenting with symptoms consistent with acute coronary syndrome (ACS).

All subject diagnoses were adjudicated by panels of certified cardiologists and emergency physicians according to the Third Universal Definition Of Myocardial Infarction - consensus guideline<sup>9</sup> endorsed by the European Society of Cardiology (ESC), the American College of Cardiology Foundation (ACCF), the American Heart Association (AHA), and the World Heart Federation (WHF). The observed AMI prevalence in this study was 13%.

The results from the study are presented using serial time intervals analyzed according to:

- the time from baseline blood draw
- the time of presentation to the emergency department

b Not detectable (< 0.01%).

Results were established using the Atellica IM Analyzer. The cutoff value, and sensitivity and specificity information should only be used as a guide when determining the appropriate cutoff value for your institution. Because sensitivity and specificity are influenced by the cutoff value, laboratories should select a cutoff value based on their specific sensitivity and specificity requirements.

### Analysis 1: Time from baseline blood draw

The results were analyzed using serial time intervals subsequent to a baseline blood draw collected for the study during the emergency department visit. The baseline blood draw for the study was collected 45 minutes (median) after the emergency department blood draw.

**Note** The initial emergency department blood draw occurred at a median of 44 minutes (interquartile range of 26–70 minutes) following patient admission.

The pooled gender results that are summarized in table 1 are based on time from the study baseline blood draw, calculated using the overall 99th percentile of 45.20 pg/mL (ng/L). Gender-specific data are presented in tables 2 and 3:

Table 1: Pooled gender results based on time from study baseline blood draw

Time-Point	Sensitivity		Specificity		Positive Predictive Value			Negative Predictive Value				
(hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Lithium-hepari	n plas	ma										
Baseline	299	84.3	79.7–88.0	2003	90.8	89.4–92.0	437	57.7	53.0-62.2	1865	97.5	96.7–98.1
≥ 0.75-< 1.5	255	90.6	86.4–93.6	1871	90.8	89.4–92.0	403	57.3	52.4-62.1	1723	98.6	97.9–99.1
≥ 1.5-< 2.5	138	92.8	87.2–96.0	1042	89.8	87.8-91.5	234	54.7	48.3-61.0	946	98.9	98.1–99.4
≥ 2.5-< 3.5	118	93.2	87.2–96.5	682	90.0	87.6-92.1	178	61.8	54.5-68.6	622	98.7	97.5–99.3
≥ 3.5-< 9	251	94.0	90.4–96.3	1120	86.9	84.8-88.7	383	61.6	56.7-66.4	988	98.5	97.5–99.1
≥ 9–24	224	92.4	88.2-95.2	885	86.6	84.1-88.6	326	63.5	58.1-68.5	783	97.8	96.6–98.6
Serum												
Baseline	294	84.7	80.1-88.4	2035	91.0	89.7–92.2	432	57.6	52.9-62.2	1897	97.6	96.8–98.2
≥ 0.75-< 1.5	252	88.1	83.5–91.5	1881	91.1	89.8-92.3	389	57.1	52.1-61.9	1744	98.3	97.6–98.8
≥ 1.5-< 2.5	134	91.8	85.9–95.4	1048	90.0	88.0-91.7	228	53.9	47.5-60.3	954	98.8	97.9–99.4
≥ 2.5-< 3.5	113	92.0	85.6-95.8	680	90.4	88.0-92.4	169	61.5	54.0-68.5	624	98.6	97.3–99.2
≥ 3.5-< 9	245	94.7	91.1–96.9	1124	87.7	85.7-89.5	370	62.7	57.7-67.5	999	98.7	97.8–99.2
≥ 9–24	225	91.6	87.2-94.5	895	87.4	85.0-89.4	319	64.6	59.2-69.6	801	97.6	96.3-98.5

a Number of samples.

b Confidence interval.

Results for females based on time from study baseline blood draw, calculated using the female-specific 99th percentile values of 34.11 pg/mL (ng/L) for plasma and 38.64 pg/mL (ng/L) for serum are summarized in table 2.

Table 2: Results for females based on time from study baseline blood draw

Time-Point	Sensitivity Point ————————————————————————————————————		Spec	Specificity		Positive Predictive Value			Negative Predictive Value			
(hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Lithium-hepari	n plas	ma										
Baseline	106	87.7	80.1–92.7	903	91.4	89.4-93.0	171	54.4	46.9-61.7	838	98.4	97.4–99.1
≥ 0.75-< 1.5	92	90.2	82.4–94.8	835	91.5	89.4–93.2	154	53.9	46.0-61.6	773	98.8	97.8–99.4
≥ 1.5-< 2.5	43	97.7	87.9–99.6	435	91.7	88.8-94.0	78	53.8	42.9-64.5	400	99.8	98.6-100.0
≥ 2.5-< 3.5	41	95.1	83.9–98.7	318	87.7	83.7–90.9	78	50.0	39.2-60.8	281	99.3	97.4–99.8
≥ 3.5-< 9	92	94.6	87.9–97.7	468	88.2	85.0-90.9	142	61.3	53.1-68.9	418	98.8	97.2-99.5
≥ 9–24	77	93.5	85.7–97.2	360	86.1	82.2-89.3	122	59.0	50.1-67.3	315	98.4	96.3–99.3
Serum												
Baseline	101	86.1	78.1–91.6	911	91.7	89.7–93.3	163	53.4	45.7-60.9	849	98.4	97.3-99.0
≥ 0.75-< 1.5	88	88.6	80.3-93.7	830	91.7	89.6-93.4	147	53.1	45.0-60.9	771	98.7	97.6-99.3
≥ 1.5-< 2.5	40	95.0	83.5–98.6	431	92.3	89.4–94.5	71	53.5	42.0-64.6	400	99.5	98.2-99.9
≥ 2.5-< 3.5	38	94.7	82.7–98.5	322	88.8	84.9-91.8	72	50.0	38.7-61.3	288	99.3	97.5–99.8
≥ 3.5-< 9	87	95.4	88.8-98.2	476	89.1	86.0-91.6	135	61.5	53.1-69.3	428	99.1	97.6–99.6
≥ 9–24	76	93.4	85.5–97.2	360	88.6	84.9–91.5	112	63.4	54.2-71.7	324	98.5	96.4–99.3

<sup>&</sup>lt;sup>a</sup> Number of samples.

Results for males based on time from study baseline blood draw, calculated using the male-specific 99th percentile values of 53.48 pg/mL (ng/L) for plasma and 53.53 pg/mL (ng/L) for serum are summarized in table 3.

Table 3: Results for males based on time from study baseline blood draw

Sensitivity Time-Point		Specificity		Positive Predictive Value			Negative Predictive Value					
(hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Lithium-hepari	n plas	ma										
Baseline	193	81.9	75.8–86.7	1100	91.1	89.3–92.6	256	61.7	55.6-67.5	1037	96.6	95.3–97.6
≥ 0.75-< 1.5	163	89.0	83.2-92.9	1036	91.0	89.1–92.6	238	60.9	54.6-69.9	961	98.1	97.1–98.8
≥ 1.5-< 2.5	95	88.4	80.4-93.4	607	89.8	87.1–92.0	146	57.5	49.4-65.3	556	98.0	96.5–98.9
≥ 2.5-< 3.5	77	90.9	82.4–95.5	364	91.2	87.9–93.7	102	68.6	59.1–76.8	339	97.9	95.8–99.0
≥ 3.5-< 9	159	93.1	88.0–96.1	652	86.8	84.0-89.2	234	63.2	56.9-69.2	577	98.1	96.6–98.9
≥ 9–24	147	91.2	85.5–94.8	525	87.4	84.3-90.0	200	67.0	60.2-73.1	472	97.2	95.3–98.4

b Confidence interval.

Sensitivity Time-Point					Positive Predictive Value			Negative Predictive Value				
(hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Serum												
Baseline	193	82.9	77.0-87.6	1124	91.2	89.4–92.7	259	61.8	55.7-67.5	1058	96.9	95.7–97.8
≥ 0.75-< 1.5	164	87.2	81.2-91.5	1051	91.2	89.4-92.8	235	60.9	54.5-66.9	980	97.9	96.7–98.6
≥ 1.5-< 2.5	94	87.2	79.0–92.5	617	89.8	87.1–91.9	145	56.6	48.4-64.3	566	97.9	96.3-98.8
≥ 2.5-< 3.5	75	90.7	82.0-95.4	358	91.3	88.0-93.8	99	68.7	59.0-77.0	334	97.9	95.7–99.0
≥ 3.5-< 9	158	93.0	88.0–96.1	648	87.8	85.1–90.1	226	65.0	58.6-71.0	580	98.1	96.6–98.9
≥ 9–24	149	89.9	84.1–93.8	535	87.9	84.8–90.4	199	67.3	60.5-73.5	485	96.9	95.0–98.1

<sup>&</sup>lt;sup>a</sup> Number of samples.

### Analysis 2: Time of presentation to the emergency department

The pooled gender results based on time of presentation to the emergency department, calculated using the overall 99th percentile of 45.20 pg/mL (ng/L), are summarized in table 4. Gender-specific data are presented in tables 5 and 6.

Table 4: Pooled gender results based on time from presentation to the emergency department

Time Deint	Sensitivity me-Point ————————————————————————————————————		Specificity		Posit Valu		edictive	Negative Predictive Value				
(hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Lithium-hepa	rin pla	sma										
0-<1.5	145	77.2	69.8-83.3	964	91.8	89.9-93.4	190	58.9	51.8-65.7	919	96.4	95.0-97.4
≥ 1.5-< 2.5	240	90.0	85.6-93.2	1625	90.6	89.1–91.9	368	58.7	53.6-63.6	1497	98.4	97.6–98.9
≥ 2.5-< 3.5	201	92.0	87.5-95.0	1369	90.6	88.9-92.0	314	58.9	53.4-64.2	1256	98.7	97.9–99.2
≥ 3.5-< 4.5	149	92.6	87.3-95.8	1080	90.9	89.1–92.5	237	58.2	51.9-64.3	992	98.9	98.0-99.4
≥ 4.5-< 6	66	97.0	89.6–99.2	461	89.2	86.0-91.7	114	56.1	47.0-64.9	413	99.5	98.3-99.9
≥ 6-<9	193	92.7	88.2-95.6	905	87.6	85.3-89.6	291	61.5	55.8-66.9	807	98.3	97.1–99.0
≥ 9-< 24	215	94.0	89.9–96.4	835	86.6	84.1-88.7	314	64.3	58.9-69.4	736	98.2	97.0–99.0
≥ 24	62	93.5	84.6–97.5	253	83.4	78.3–87.5	100	58.0	48.2-67.2	215	98.1	95.3-99.3
Serum												
0-<1.5	140	78.6	71.1–84.6	980	92.1	90.3-93.7	186	59.1	52.0-65.9	934	96.8	95.4-97.7
≥ 1.5-< 2.5	239	87.9	83.1–91.4	1646	90.9	89.5–92.2	358	58.7	53.5-63.6	1527	98.1	97.3–98.7
≥ 2.5-< 3.5	195	90.8	85.9-94.1	1384	90.8	89.1–92.2	305	58.0	52.4-63.4	1274	98.6	97.8–99.1
≥ 3.5-< 4.5	147	91.2	85.5-94.8	1083	91.0	89.2-92.6	232	57.8	51.3-63.9	998	98.7	97.8–99.2
≥ 4.5-< 6	64	96.9	89.3–99.1	454	89.6	86.5-92.1	109	56.9	47.5–65.8	409	99.5	98.2-99.9
≥ 6-<9	185	93.5	89.0–96.3	904	88.7	86.5-90.6	275	62.9	57.1–68.4	814	98.5	97.4–99.2

<sup>&</sup>lt;sup>b</sup> Confidence interval.

Sensitivity Time-Point ————		Specificity			Positive Predictive Value			Negative Predictive Value				
(hours)	Nª	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
≥ 9-< 24	213	93.9	89.8–96.4	841	86.7	84.2-88.8	312	64.1	58.6-69.2	742	98.2	97.0–99.0
≥ 24	63	90.5	80.7-95.6	255	85.5	80.6-89.3	94	60.6	50.5-69.9	224	97.3	94.3–98.8

<sup>&</sup>lt;sup>a</sup> Number of samples.

Results for females based on time of presentation to the emergency department, calculated using the female-specific 99th percentile of 34.11 pg/mL (ng/L) for plasma and 38.64 pg/mL (ng/L) for serum are summarized in table 5.

Table 5: Results for females based on time of presentation to the emergency department

Time-Point	Sensitivity t ———————————————————————————————————		Spec	Specificity		Posit Value	ive Pre	dictive	Negative Predictive Value			
(hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Lithium-hepa	rin pl	asma										
0-<1.5	45	84.4	71.2–92.3	401	93.3	90.4-95.3	64	59.4	47.1–70.5	382	98.2	96.3–99.1
≥ 1.5-< 2.5	79	89.9	81.3-94.8	720	91.7	89.4–93.5	130	54.6	46.0-62.9	669	98.8	97.7–99.4
≥ 2.5-< 3.5	73	94.5	86.7–97.8	621	91.6	89.2–93.6	121	57.0	48.1–65.5	573	99.3	98.2–99.7
≥ 3.5-< 4.5	50	94.0	83.8–97.9	487	89.5	86.5-91.9	98	48.0	38.3–57.7	439	99.3	98.0–99.8
≥ 4.5-< 6	26	96.2	81.1–99.3	238	87.0	82.1–90.7	56	44.6	32.4–57.6	208	99.5	97.3–99.9
≥ 6-<9	69	94.2	86.0-97.7	374	88.0	84.3-90.9	110	59.1	49.7–67.8	333	98.8	97.0–99.5
≥ 9-< 24	74	94.6	86.9–97.9	342	87.4	83.5-90.5	113	61.9	52.7-70.4	303	98.7	96.7–99.5
≥ 24	27	96.3	81.7–99.3	110	80.9	72.6-87.2	47	55.3	41.2-68.6	90	98.9	94.0-99.8
Serum												
0-<1.5	42	81.0	66.7–90.0	407	93.4	90.5-95.4	60	56.7	44.1–68.4	389	97.9	96.0–99.0
≥ 1.5-< 2.5	77	89.6	80.8-94.6	721	91.8	89.6–93.6	127	54.3	45.7–62.7	671	98.8	97.7–99.4
≥ 2.5-< 3.5	67	92.5	83.7–96.8	619	92.4	90.0-94.2	109	56.9	47.5–65.8	577	99.1	98.0–99.6
≥ 3.5-< 4.5	48	93.8	83.2-97.9	484	90.3	87.3-92.6	92	48.9	38.9–59.0	440	99.3	98.0–99.8
≥ 4.5-< 6	26	96.2	81.1–99.3	236	87.7	82.9-91.3	54	46.3	33.7–59.4	208	99.5	97.3–99.9
≥ 6-<9	63	95.2	86.9–98.4	378	88.6	85.0-91.4	103	58.3	48.6-67.3	338	99.1	97.4–99.7
≥ 9-< 24	73	94.5	86.7–97.8	342	89.5	85.8-92.3	105	65.7	56.2-74.1	310	98.7	96.7–99.5
≥ 24	26	96.2	81.1–99.3	111	82.9	74.8–88.8	44	56.8	42.2-70.3	93	98.9	94.2–99.8

<sup>&</sup>lt;sup>a</sup> Number of samples.

b Confidence interval.

b Confidence interval.

Results for males based on time of presentation to the emergency department, calculated using the male-specific 99th percentile of 53.48 pg/mL (ng/L) for plasma and 53.53 pg/mL (ng/L) for serum, are summarized in table 6.

Table 6: Results for males based on time of presentation to the emergency department

T. D.	Sensitivity		Specificity		Posit Valu		dictive	Negative Predictive Value				
Time-Point (hours)	Na	%	95% CI <sup>b</sup>	N	%	95% CI	N	%	95% CI	N	%	95% CI
Lithium-hepa	rin pla	sma										
0-<1.5	100	75.0	65.7-82.5	563	91.5	88.9–93.5	123	61.0	52.1-69.1	540	95.4	93.3–96.8
≥ 1.5-< 2.5	161	87.6	81.6-91.8	905	91.2	89.1–92.8	221	63.8	57.3-69.9	845	97.6	96.4–98.5
≥ 2.5-< 3.5	128	89.8	83.4-94.0	748	90.0	87.6-91.9	190	60.5	53.4-67.2	686	98.1	96.8–98.9
≥ 3.5-< 4.5	99	90.9	83.6-95.1	593	92.4	90.0-94.3	136	66.2	57.9-73.6	556	98.4	97.0–99.1
≥ 4.5-< 6	40	92.5	80.1–97.4	223	90.6	86.0-93.8	58	63.8	50.9-74.9	205	98.5	95.8–99.5
≥ 6-<9	124	91.1	84.8-95.0	531	88.5	85.5-91.0	174	64.9	57.6-71.6	481	97.7	96.0–98.7
≥ 9-< 24	141	93.6	88.3–96.6	493	85.6	82.2-88.4	203	65.0	58.2-71.3	431	97.9	96.1–98.9
≥ 24	35	91.4	77.6–97.0	143	88.1	81.8-92.4	49	65.3	51.3-77.1	129	97.7	93.4–99.2
Serum												
0-<1.5	98	77.6	68.3-84.7	573	91.6	89.1–93.6	124	61.3	52.5-69.4	547	96.0	94.0-97.3
≥ 1.5-< 2.5	162	86.4	80.3-90.9	925	91.5	89.5–93.1	219	63.9	57.4-70.0	868	97.5	96.2-98.3
≥ 2.5-< 3.5	128	87.5	80.7–92.2	765	89.9	87.6-91.9	189	59.3	52.1-66.0	704	97.7	96.3–98.6
≥ 3.5-< 4.5	99	88.9	81.2-93.7	599	92.7	90.3-94.5	133	66.2	57.8-73.7	565	98.1	96.6–98.9
≥ 4.5-< 6	38	94.7	82.7-98.5	218	90.8	86.3-94.0	56	64.3	51.2-75.5	200	99.0	96.4–99.7
≥ 6-<9	122	91.0	84.6-94.9	526	89.9	87.1–92.2	164	67.7	60.2-74.4	484	97.7	96.0–98.7
≥ 9-< 24	140	93.6	88.2-96.6	499	85.6	82.2-88.4	203	64.5	57.7–70.8	436	97.9	96.1–98.9
≥ 24	37	86.5	72.0-94.1	144	88.9	82.7-93.0	48	66.7	52.5-78.3	133	96.2	91.5–98.4

<sup>&</sup>lt;sup>a</sup> Number of samples.

#### **Elevated Tnl Values in Patients Without AMI**

There are conditions other than AMI that are known to cause myocardial injury and elevated TnI values. 9,11–14,27-34

The Atellica IM TnIH clinical trial enrolled all patients presenting to the emergency department with symptoms consistent with ACS. Some of these patients had an acute or chronic condition other than AMI.

In the clinical trial, 11% of patients without an AMI diagnosis had at least 1 Atellica IM TnIH test result above the 99th percentile (>45.20~pg/mL~(ng/L)) on 1 or more serial draws. 88% of these patients were found to have 1 or more of the following conditions:

#### **Cardiac conditions**

- Angina
- Atrial fibrillation
- Cardiomyopathy

<sup>&</sup>lt;sup>b</sup> Confidence interval.

- · Coronary artery disease
- Heart failure
- Hypertensive urgency
- Pericarditis
- Recent cardiac intervention
- Severe valvular heart disease
- Tachycardia

#### **Non-cardiac conditions**

- Chronic lung disease
- Cardiac contusion related to a traumatic injury
- Renal failure
- Pneumonia
- Pulmonary embolism
- Shock
- Systemic sclerosis

### **Precision**

Precision was determined in accordance with CLSI Document EP05-A3.<sup>35</sup> Samples were assayed on an Atellica CI Analyzer in duplicate in 2 runs per day for 20 days. The following results are representative of the performance of the assay:

			Repeatability		Within-Laborat	ory Precision
Sample Type	Nª	Mean (pg/mL; ng/L)	SD <sup>b</sup> (pg/mL; ng/L)	CV <sup>c</sup> (%)	SD (pg/mL; ng/L)	CV (%)
Serum 1	80	13.73	0.336	2.4	0.417	3.0
Serum 2	80	129.63	1.535	1.2	2.540	2.0
Serum 3	80	1262.15	14.471	1.1	19.601	1.6
Serum 4	80	12345.45	91.546	0.7	158.205	1.3
Serum 5	80	23197.12	176.231	0.8	396.901	1.7
Plasma 1	80	15.83	0.698	4.4	0.810	5.1
Plasma 2	80	160.58	2.429	1.5	3.306	2.1
Plasma 3	80	1493.84	23.895	1.6	27.337	1.8
Plasma 4	80	13068.63	116.135	0.9	205.890	1.6
Plasma 5	80	21143.05	165.405	0.8	383.217	1.8

<sup>&</sup>lt;sup>a</sup> Number of samples tested.

Assay results obtained at individual laboratories may vary from the data presented.

b Standard deviation.

<sup>&</sup>lt;sup>c</sup> Coefficient of variation.

## **Assay Comparison**

Assay comparison was determined with the weighted Deming regression model in accordance with CLSI Document EP09-A3.<sup>36</sup>

Agreement of the assays may vary depending on the study design, comparative assay, and population tested.

Specimen Type	Comparative Assay (x)	Regression Equation	Sample Interval	Nª	r <sup>b</sup>
Lithium heparin plasma	Atellica IM TnIH on Atellica IM Analyzer	y = 1.00x + 0.04 pg/mL (ng/L)	2.66-23395.95 pg/mL (ng/L)	170	0.996

<sup>&</sup>lt;sup>a</sup> Number of samples tested.

### **Interferences**

Interference testing was performed in accordance with CLSI Document EP07-A2.<sup>25</sup>

Testing was performed with both human serum and lithium-heparin plasma samples, with troponin concentrations in the ranges of 20-60 pg/mL (ng/L) and 1000-2000 pg/mL (ng/L). The following drugs were added to the samples at the concentrations indicated, and were evaluated for potential interference using the Atellica IM TnIH assay. The results demonstrated a  $\leq 10\%$  interference from each drug.

	Low or Therapeution	c Concentration	High or Toxic Co	ncentration
Potential Interferents	Common units	SI units	Common units	SI units
Abciximab	5 μg/mL	N/Aª	40 μg/mL	N/A
Acetaminophen	20 μg/mL	133 μmol/L	200 μg/mL	1324 μmol/L
Acetylsalicylic acid	261 μg/mL	1.45 mmol/L	652 μg/mL	3.62 mmol/L
Allopurinol	13 μg/mL	92 μmol/L	40 μg/mL	294 μmol/L
Amiodarone	1.8 μg/mL	2.6 μmol/L	6.1 μg/mL	8.92 μmol/L
Ampicilin	10 μg/mL	29.1 μmol/L	53 μg/mL	152 μmol/L
Ascorbic acid	12 μg/mL	68.5 μmol/L	60 μg/mL	342 μmol/L
Atenolol	1.1 μg/mL	4.14 µmol/L	10 μg/mL	37.6 μmol/L
Caffeine	12 μg/mL	64.4 µmol/L	60 μg/mL	308 µmol/L
Captropril	1.0 μg/mL	4.6 μmol/L	5.0 μg/mL	23 μmol/L
Cefoxitin	120 μg/mL	281 μmol/L	660 μg/mL	1546 μmol/L
Cinnarizine	200 ng/mL	542 nmol/L	400 ng/mL	1084 nmol/L
Clopidogrel	37.5 μg/mL	116 µmol/L	75 μg/mL	233 μmol/L
Cocaine	0.1 μg/mL	0.33 μmol/L	10 μg/mL	33 μmol/L
Digoxin	1.4 ng/mL	1.8 nmol/L	6.1 ng/mL	7.8 nmol/L
Digitoxin	30 ng/mL	39 nmol/L	60 ng/mL	78 nmol/L
Diltiazem	0.2 μg/mL	0.55 μmol/L	6.2 μg/mL	15 μmol/L

b Correlation coefficient.

	Low or Therapeut	tic Concentration	High or Toxic Co	ncentration
Potential Interferents	Common units	SI units	Common units	SI units
Disopyramide	3.5 μg/mL	10.4 μmol/L	10 μg/mL	29.5 μmol/L
Dopamine	0.3 μg/mL	1.96 μmol/L	0.9 μg/mL	5.87 µmol/L
Doxycycline	10.0 μg/mL	22.5 μmol/L	30 μg/mL	67.5 µmol/L
Erythromycin	11 μg/mL	14.96 µmol/L	60 μg/mL	81.6 µmol/L
Furosemide	20 μg/mL	60.4 μmol/L	60 μg/mL	181 µmol/L
Ibuprofen	40 μg/mL	194.3 µmol/L	500 μg/mL	2425 μmol/L
Isosorbide dinitrate	50 ng/mL	212 nmol/L	150 ng/mL	636 nmol/L
Lisinopril	0.10 μg/mL	0.25 μmol/L	0.30 μg/mL	0.74 µmol/L
Lovastatin	40 ng/mL	95 nmol/L	80 ng/mL	191 nmol/L
Low MW heparin	6.75 U/mL	N/A	30 U/mL	N/A
Methotrexate	546 μg/mL	1.2 mmol/L	910 μg/mL	2.0 mmol/L
Methyldopa	4.2 μg/mL	20.12 μmol/L	15 μg/mL	71 µmol/L
Methylprednisolone	N/A	N/A	40 μg/mL	107 µmol/L
Mexiletine	1.3 μg/mL	7 μmol/L	4.0 μg/mL	22.3 µmol/L
Nicotine	37 ng/mL	0.23 μmol/L	1000 ng/mL	6.2 µmol/L
Nifedipine	125 ng/mL	362 nmol/L	400 ng/mL	1156 nmol/L
Nitrofurantoin	2.0 μg/mL	8.4 μmol/L	4.0 μg/mL	16.8 µmol/L
Nitroglycerine	7.5 ng/mL	33 nmol/L	160 ng/mL	704 nmol/L
Phenobarbital	24 μg/mL	107.6 μmol/L	97 μg/mL	431 µmol/L
Phenytoin	12 μg/mL	49.5 μmol/L	50 μg/mL	198 µmol/L
Primidone	10.5 μg/mL	48.2 μmol/L	40 μg/mL	183 µmol/L
Propranolol	0.50 μg/mL	1.94 µmol/L	2.0 μg/mL	7.71 µmol/L
Quinidine	3.7 μg/mL	11.56 μmol/L	12 μg/mL	37 μmol/L
Simvastatin	16 μg/mL	38 μmol/L	32 μg/mL	76 µmol/L
Theophylline	12 μg/mL	69.4 μmol/L	40 μg/mL	222 µmol/L
Thyroxine	0.08 μg/mL	0.11 μmol/L	1.01 μg/mL	1.30 µmol/L
Tissue plasminogen activator (TPA)	1.15 μg/mL	N/A	2.3 μg/mL	N/A
Trimethoprim	12 μg/mL	43 µmol/L	40 μg/mL	138 µmol/L
Verapamil	0.33 μg/mL	0.72 μmol/L	2.0 μg/mL	4.4 µmol/L
Warfarin	2.0 μg/mL	6.6 µmol/L	10 μg/mL	32.5 μmol/L

<sup>&</sup>lt;sup>a</sup> Not applicable.

Results were established using the Atellica IM Analyzer. Assay results obtained at individual laboratories may vary from the data presented.

#### Hemolysis, Icterus, Lipemia (HIL), and Other Interferences

Potential interference in the Atellica IM TnIH assay from the compounds listed below is designed to be  $\leq 10\%$ .

Specimens that are	Demonstrate ≤ 10% change in results up to
Hemolyzed	500 mg/dL of hemoglobin
Lipemic	2000 mg/dL of triglycerides
Icteric	40 mg/dL of conjugated bilirubin
Icteric	60 mg/dL of unconjugated bilirubin

Specimens that contain	Demonstrate ≤ 10% change in results up to	
Biotin	3500 ng/mL of biotin	
Cholesterol	500 mg/dL of cholesterol	
Protein Albumin	6 g/dL	
Protein Gamma Globulin	2.5 g/dL	
Total Protein	12 g/dL	

Results were established using the Atellica IM Analyzer. Assay results obtained at individual laboratories may vary from the data presented.

## Linearity

The Atellica IM TnIH assay is linear from 2.50–25,000.00 pg/mL (ng/L). Linearity was evaluated using the Atellica CI Analyzer according to the CLSI Document EP06-ed2.<sup>37</sup> Serum and lithium-heparin plasma samples were used to make pools at 3 different cTNI ranges. The dilution series were made by mixing high and low concentration samples. The resulting sample mixtures were tested with the Atellica IM TnIH assay.

# **High-Dose Hook Effect**

High cTnI concentrations can cause a paradoxical decrease in the RLUs (high-dose hook effect). In this assay, patient samples with cTnI concentrations as high as 500,000 pg/mL (ng/L) will report > 25,000.00 pg/mL (ng/L).

### Standardization

The Atellica IM TnIH assay standardization is traceable to an internal standard manufactured using human heart homogenate. Assigned values for calibrators are traceable to this standardization.

## **Technical Assistance**

For customer support, contact your local technical support provider or distributor. siemens-healthineers.com

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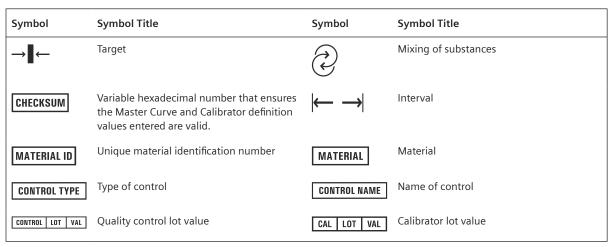
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# **Definition of Symbols**

The following symbols may appear on the product labeling:

Symbol	Symbol Title	Symbol	Symbol Title
	Manufacturer	EC REP	Authorized representative in the European Community
	Use-by date	CH REP	Authorized representative in Switzerland
REF	Catalog number	LOT	Batch code
(i)	Consult Instructions for Use	Σ	Contains sufficient for <n> tests</n>
<b>i</b>	Internet URL address to access the electronic instructions for use	Rev. XX	Version of Instructions for Use
IVD	In vitro diagnostic medical device	Rev.	Revision
RxOnly	Prescription device (US only)	UDI	Unique Device Identifier
<b>C</b> € xxxx	CE Marking with Notified Body	CE	CE Marking
1	Temperature limit	*	Keep away from sunlight
1	Upper limit of temperature	1	Lower limit of temperature
2	Do not re-use	(PCC)	Do not freeze
(A)	Recycle	<u>††</u>	This way up
8	Biological risks	$\triangle$	Caution
UNITS C	Common Units	UNITS SI	International System of Units
YYYY-MM-DD	Date format (year-month-day)	YYYY-MM	Date format (year-month)
	Document face up <sup>a</sup>		Handheld barcode scanner



a Indicates Assay-eNote

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